



Raman Spectroscopy for the creation of M412 Intermediate

Govindaswamy.R¹

Research Scholar

*Research & Development Centre,
Bharathiar University,
Coimbatore, Tamil Nadu.*

Dr.O.N.Balasundaram²

Associate Professor,

*Department of Physics,
PSG College of Arts and Science,
(Affiliated to Bharathiar University)
Coimbatore, Tamil Nadu.*

Abstract

To dop the M intermediate, bcbR crystals were related to both photo-excitation and dehydration resonance. In this study, the presence of the M intermediate is exploited as a cue to search for traces of a photocycle using raman spectroscopy of single hydrated bcbR crystals were studied and resonance before using bacteriorhodopsin

4482

Keywords: photo-excitation, Raman Spectroscopy, resonance

DOI Number:10.48047/nq.2022.20.22.NQ10450 **NeuroQuantology**2022;20(22):4482-4488

Introduction

A fraction of an incident radiation beam is scattered in every direction by the species present after passing through a medium. In 1928, C.V. Raman made the observation that a small percentage of the radiation dispersed by specific molecules had a wavelength that is different from the incident beam. The chemical composition of the molecules that cause the scattering determines how the wavelength shifts (1). The energy of the vibration that caused the scattering corresponds to the energy difference between incident radiation and scattered radiation. Although the molecule's vibrational state must change for one to be able to see an infrared absorption spectrum. Resonant energy from the incident laser Raman spectroscopy determines how much energy a molecule has by analysing the chromophore's electronic transitions. Drawings of the resonant Raman and Rayleigh scattering processes are shown,

which were taken from Ferraro et al. (2). The entering laser wavelength in resonance Rayleigh and Raman spectroscopy is chosen to match the energy of an electronic transition in a chromophore. The laser's energy is sufficient to provide the molecule the energy it needs to induce an excited state. This chromophore's Raman bands are selectively enhanced³ by a factor of 10^3 - 10^5 .

This chromophore's vibration produces Raman bands that are specially multiplied by a factor (2) of 10^3 - 10^5 . This significant improvement is crucial for understanding biological systems because it allows researchers to examine chromophore changes without being hindered by the protein's many amino acid residues.

These experiments focus on the retinal protein bacteriorhodopsin, it has an absorption spectrum observable to the naked eye with at least two distinct bands between 250 and 700 nm. Theoretically, the



dark-adapted species' Raman spectra, acquired using an 830 nm laser, will have bands corresponding to both the 13-cis and all-trans isomers. A protein that has only recently been light-adapted will only exhibit bands generated by the all-trans retinal species when it is analysed using the same 830 nm laser. However, using an incoming laser whose wavelength is in the retinals' absorption band, both the parent bR's resonance-enhanced retinal spectrum and any intermediate photo products created by photochemistry ought to be distinct.

Water is transparent in Raman spectroscopy because of its small Raman scattering cross-section, when it comes to the comprehensive analysis of hydrated biological systems, this is one of the most significant advantages that Raman spectroscopy offers over infrared spectroscopy. The molecule is already polarised, hence as a result of this dipole, there are no Raman bands produced, especially when the laser's wavelength is extremely far from the molecule's vacuum UV absorption. Contrarily, water produces strong IR bands that frequently overlap, preventing the detection of numerous protein vibrations in the 2000–1300 cm^{-1} range.

The chromophore spends the majority of the photocycle in the 13-cis form. The M intermediate stands out because it is the only one where the Schiff base is deprotonated, despite retinal being in the 13-cis configuration in multiple photocycle intermediates. The fingerprint region's (13-cis configuration) 1191 cm^{-1} band will also develop, and the frequency of C=C stretching will change from 1528 to 1566 cm^{-1} .

The study here aims to compile Raman spectra of the bcbR crystal at the incident wavelengths of 830 nm and 514 nm and to contrast them with the information data collected for bR in water membrane native solutions under equivalent experimental conditions. The spectra of the light-adapted bR is gathered using an 830 nm Raman laser. Since the incoming wavelength is far from the chromophore's 570 nm absorption maximum, only the unphotolyzed bR

species should scatter at this wavelength. To excite the material and acquire the spectra of any likely combination of intermediates, it employs a 514 nm Raman incident laser. New bands should emerge close to 1566 cm^{-1} and between 1167 and 1201 cm^{-1} if the crystal goes through a photocycle. If the Schiff base deprotonates at any time while the M intermediate is being created, the C=N stretch from 1640 to 1620 cm^{-1} should likewise change.

To rule out resonance amplification as the source of any observable extra bands following stimulation with a 514 nm laser, it is necessary to do a series of experiments, comparison of the strengths of the most prominent bR bands to those that define the M state is studied. The 1566 cm^{-1} band and 1525 cm^{-1} band are the two most pronounced bands. When the wavelength of the incoming Raman laser was changed from 830 to 514 nm, one would expect resonance enhancement to result in an increase in all bands. However, if the M intermediate is produced by the 514 nm laser as well as photolyzing bR, As the 514 nm laser's power is increased, the ratio of these two bands should change. If the incoming laser is just creating resonance amplification of all bands and isn't producing M, we would assume that when the 514 nm laser intensity increases, the ratio of the two bands will remain constant. If the 514 nm incoming laser also causes the M intermediate to develop in the crystals, then raising the power of the 514 nm laser should result in an increase in the relative quantity of M intermediate, which would be represented by an increase in the ratio of the 1566 cm^{-1} to 1525 cm^{-1} intensities.

We check that any extra bands obtained under 514 nm laser stimulation are not resonance amplified by comparing the intensities of the majority of the bR bands to those characterising the M state with increasing 514 nm laser power. The most noticeable bands are the 1566 cm^{-1} band and 1525 cm^{-1} . One would anticipate that resonance enhancement would lead to an increase in all bands when the incoming Raman laser wavelength was changed from 830 to 514 nm. We should observe a shift in



the ratio of these two bands as the 514 nm laser's power is increased, though, if it also photolyzes bR, which also produces the M intermediate. If the increase in 514 nm laser intensity is responsible only for resonance amplification of all bands and not the development of M, then the ratio between the two bands should remain constant. Increasing the power of the 514 nm incident laser would be expected to increase the relative amount of M intermediate, as shown by an increase in the ratio of the 1566 cm^{-1} to the 1525 cm^{-1} intensities, if this is indeed the case and the 514 nm laser is also responsible for causing the formation of the M intermediate in the crystals.

Materials and methods

Growth of bcbR Crystals and Washing from their Detergent Matrix

Purification of the bR samples in accordance with accepted procedures using a bacterial cell line called *Halobacterium salinarium*. bR bicelle crystals in the form of diamonds were produced. A 40% (by weight) bicellular solution in water was made by combining the two detergents, DMPC and CHAPSO, in a 2:8:1 molar ratio. The bicellular solution and the 10 mg/mL bR solution were stored in separate vials that were kept on ice. These two vials were repeatedly pipetted on ice to create a 4:1 protein solution to bicellular solution combination that contained 8% bicelles and 8.0 mg/mL bR. 6.0 M 1, 6-hexanediol and 4 M sodium phosphate monobasic monohydrate ($\text{NaH}_2\text{PO}_4 \cdot \text{H}_2\text{O}$) were combined in a 28:6:1:1.4 volume ratio to produce a precipitant well solution. On a glass coverslip with a 22 mm diameter, an aliquot of 6 mL of the protein/bicelle solution, 2.5 mL of the well solution, and 1 mL of the 2.5% *D*-octylglucoside solution were pipetted gently together. The coverslips were placed within a crystallisation well that held 1 mL of the precipitant solution, was vacuum grease-sealed, and was maintained at 37 °C during crystallisation. The initially freely mixed suspension eventually gelled during the course of the next two weeks due to balance between the saturated solution below and the suspended solution above. Samples were frequently checked

throughout this time. After that, the slides were inspected under an optical microscope to look for signs of crystallisation. Deionized distilled water was used to repeatedly flush the crystal detergent matrix in micro centrifuge tubes to produce crystals. The white detergent particles were easy to remove since they floated in the supernatant, but the denser crystals instantly dropped at the bottom of the solution. Using this method, the detergent was removed from the crystals without affecting their quality. A concentrated drop of the crystal suspension was applied to a glass cover slip and then washed. That had been coated with aluminium. The glass cover slip was then allowed to air dry until above the drop, most of the water evaporated, locking the crystal into place on the surface. The glass slide was moistened with a single drop after the crystal had dried in order to produce the visible or Raman spectrum.

Single-Crystal Raman Spectroscopy

Only one crystal A Raman spectrometer was used to gather Raman spectra. To get the native film or single crystal's spectrum, Raman spectra were collected throughout by completely immersing the objective in water. The programme employed a silicon wafer's strong Raman band at 520 cm^{-1} for automated wave length calibration. All specified *sx-y* data were transmitted to the Origin Pro 7.5 software after being imported for baseline correction.

Result and discussion

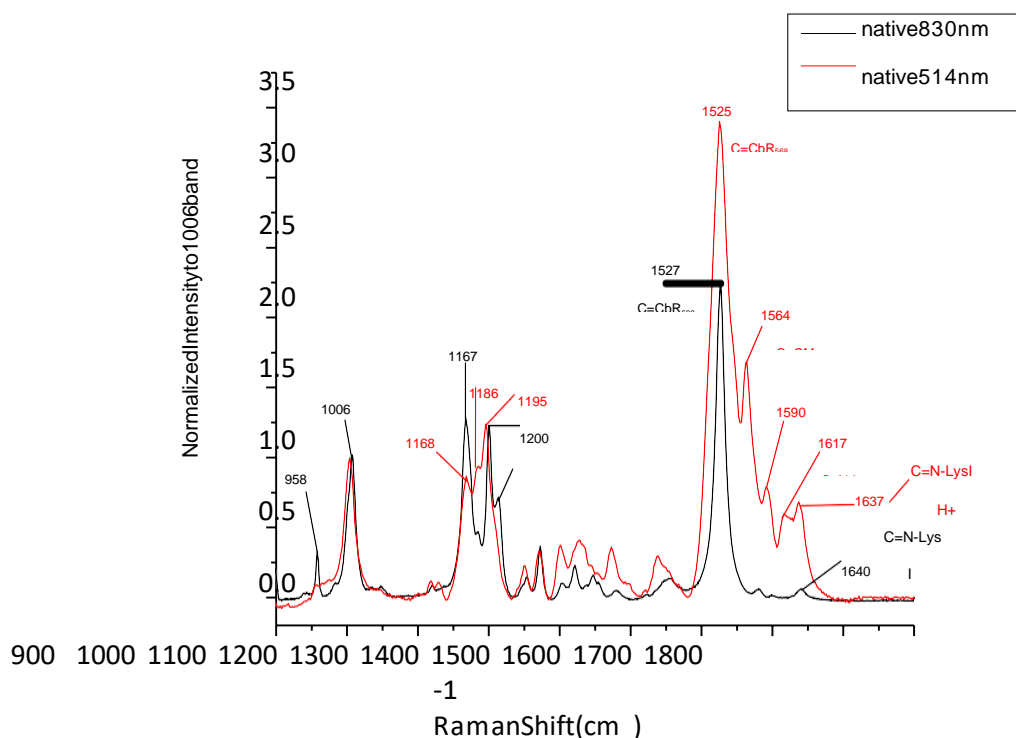
A hydrated native bR film is known to go through a photocycle. The first experiment used an 830 nm incident laser to compare the spectra of the all-trans, light-adapted bR568 film we recorded with previously reported data. Additionally, we show that the 514 nm incoming Raman laser can produce photo intermediates, which can be recognised by looking at the M intermediate Raman spectrum, in a distinct experiment. The generation of M was confirmed by a power dependency experiment with a 514nm incoming laser that intensity of the M C=C band was compared to that under non-photolytic conditions. A rise in the M/bR ratio would suggest that the M



intermediate population is also increasing as a result of the 514 nm laser, while a decline in the ratio would indicate that resonance

enhancement is the only reason utilising the 514 nm laser increases band intensities.

Single-Crystal Raman Spectroscopy of Hydrated Native bR Film at 514 nm and 830 nm



4485

Figure 1: A hydrated native bR568 film's ground state (in black) and excited state (in red) Raman spectra

The figure shows incident lasers with two distinct wavelengths —830 nm (black) and 514 nm—that irradiated a hydrated native film of bR568 to produce Raman spectra (red). Since the 514 nm laser is within the range of the retina's absorption, one would anticipate that additionally to band resonance enhancement that absorb at this wavelength, this laser can simultaneously initiate the photocycle and provide a Raman spectra of the resulting intermediates.

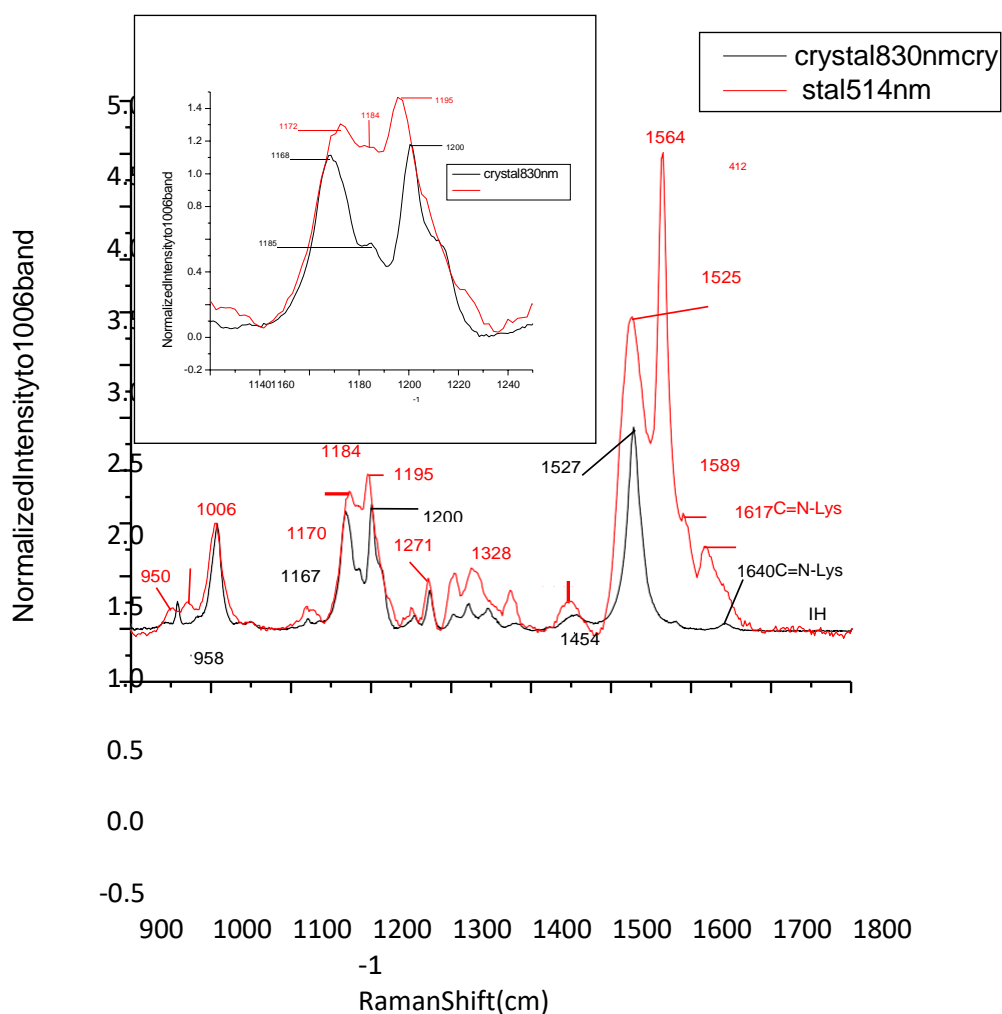
Valley between 1168 cm⁻¹ and 1195 cm⁻¹ in fingerprint area narrows when input laser wavelength is shifted from 830 nm to 514 nm. This region has been shown to be sensitive to the chromophore's isomeric conformation by time-resolved Raman spectroscopy (3),(4),(17),(15),(18).The widening of the bands due to 514 nm in this fingerprint region a change in the retina's isomeric structure to a greater population of 13-cis retinal may be indicated by raman



laser irradiation. Four picoseconds or so after excitation, retina converts to a 13-cis shape, which persists during the K, L, and M phases of the cycle (19). Therefore, the development of the 13-cis-containing photocycle intermediates of retinal is supported more by the growth of the 1186cm⁻¹ and in this area while being bathed in 514 nm light. Under 514 nm incident Raman laser, a new band at

1617cm⁻¹ emerges in the Schiff base area. This closely matches the shift of 1620 cm⁻¹ caused by the deprotonated C=N stretch (5),(6),(8),(10-12). A second band at 1637 cm⁻¹ might be a protonated Schiff base, C=N-H⁺, that is still present. The finger print region demonstrates the distinction between the 514 nm laser in the 1186 cm⁻¹ 113 cis band and the 830 nm laser with a Raman incident (black) (red).

Single-Crystal Raman Spectroscopy of Single Hydrated bcbR Crystal at 514 nm and 830 nm



4486

Figure 2: A single hydrated bicelle crystal's natural bR Raman spectrum, both unphotolyzed (in black) and photolyzed (in red), are shown.

The image displays a single hydrated bicelle crystal's Raman spectra after being exposed to input laser wavelengths of 830 nm (black) and 514 nm (white) (red). If a photocycle can be conducted by the crystal, the 514 nm

laser, which is within the retina's absorption, may be able to trigger the photocycle while sending out the spectrum of a mix of intermediaries. We can verify this by running the power dependence



experiment once more using the hydrated crystals.

The fingerprint region valley between the 1170 cm^{-1} and 1195 cm^{-1} when the input laser wavelength is altered from 830 nm to 514 nm, the bands for the hydrated bicelle crystal grow shallower. Bands intensify in the fingerprint zone as a result of 514 nm. After being exposed to a Raman laser, the retina's isomeric configuration may have shifted, has a higher population of 13-cis retinal, which is the isomer seen in the K, L, and M photocycle states (9),(11),(13),(14),(16),(17). Therefore, more proof that the retina's 13-cis-containing photocycle intermediates were formed comes from the creation of the wide

1184 cm^{-1} band in this area when exposed to 514 nm light. Under a 514 nm incident Raman laser, a new band at 1617 cm^{-1} develops in the Schiff base area. The shift of 1620 cm^{-1} brought on by the deprotonated C=N stretch is precisely matched by this. (5),(6),(8),(10-12), and was additionally noticed for the hydrated native bR film in Figure 1.

Since the 514 nm incident laser falls within the retinal chromophore's absorption band, as was already mentioned, one of the challenges in using it in these experiments is that the development of the new bands might be caused by resonance enhancement rather than the creation of the M intermediate of the photocycle.

4487

Effect of Increasing the 514 nm Laser Power on the Raman Spectrum of a Single Hydrated bcbR Crystal

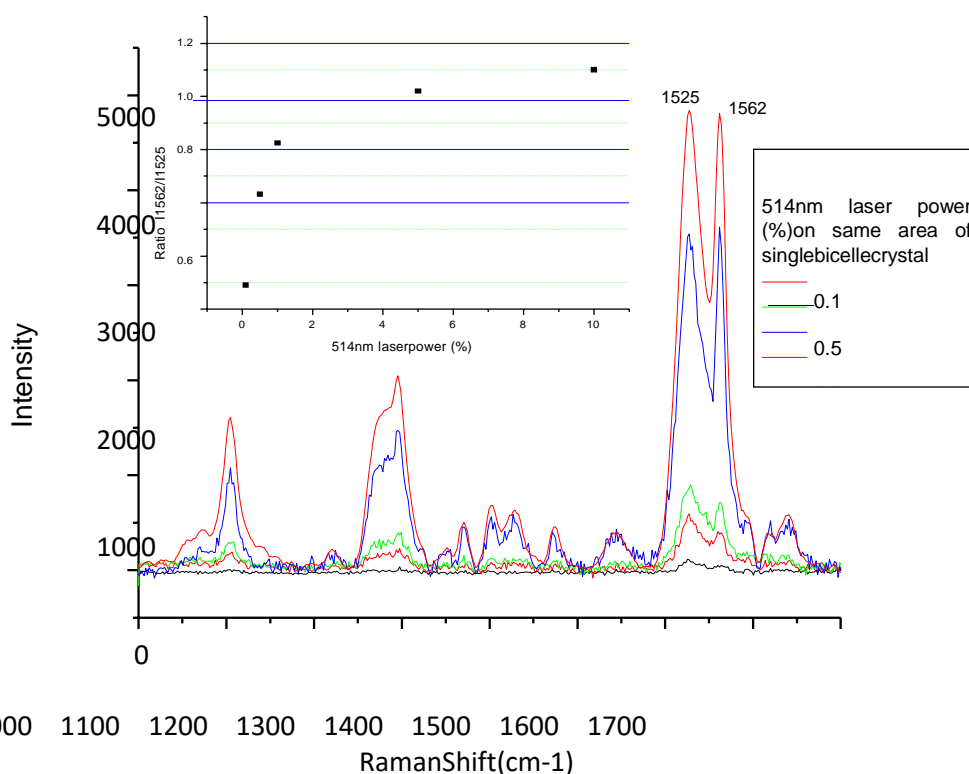


Figure 3: The Raman spectra of a single hydrated bicelle crystal vary in intensity as the incoming 514 nm laser power increases.

As seen in this picture, the strength of the 514 nm laser has an impact on the Raman spectra of a single hydrated bcbR crystal. This image's inset shows a plot of the 514 nm laser's output power against the ratio of

the 1562 cm^{-1} band to the 1525 cm^{-1} band's intensity. The rise in this ratio indicates that the M population should increase if the 514 nm laser power is raised.



Conclusion

In a single hydrated bcbR crystal, this study provides the first proof of a photocycle's existence. This is significant because it shows that this crystal form is still capable of undergoing the conformational changes brought on by proton translocation and is not stuck in a configuration that cannot be changed.

References

1. Skoog, D.A.; Leary, J.J. Raman Spectroscopy. In *Principles of Instrumental Analysis*; 4th ed.; Saunders College Publishing: Orlando, 1992; pp 296.
2. Ferraro, J.R.; Brown, K.W.; Brown, C. W. *Introductory Raman Spectroscopy*, 2nd ed.; Academic Press: Amsterdam, 2003.
3. Oesterhelt, D.; Stoekenius, W. *Nature (London), New Biology* **1971**, 233, 149.
4. Harbison, G. S.; Smith, S. O.; Pardo, J. A.; Winkel, C.; Lugtenburg, J.; Herzfeld, J.; Mathies, R.; Griffin, R. *Proceedings of the National Academy of Sciences* **1984**, 81, 1706.
5. Heyde, M.E.; Gill, D.; Kilponen, R.G.; Rimai, L. *Journal of the American Chemical Society* **1971**, 93, 6776.
6. Lewis, A.; Spoonhower, J.; Bogomolni, R.A.; Lozier, R.H.; Stoekenius, W. *Proc. Natl. Acad. Sci. U.S.A.* **1974**, 71, 4462.
8. Turner, J.; Campion, A.; El-Sayed, M. A. *Proceedings of the National Academy of Sciences of the United States of America* **1977**, 74, 5212.
9. Althaus, T.; Eisfeld, W.; Lohrmann, R.; Stockburger, M. *Israel Journal of Chemistry* **1995**, 35, 227.
10. Spiro, T. G.; Editor *Biological Applications of Raman Spectroscopy, Vol.2: Resonance Raman Spectra of Polyenes and Aromatics*, 1987.
11. Aton, B.; Doukas, A.G.; Callender, R.H.; Becher, B.; Ebrey, T.G.
12. *Biochemistry* **1977**, 16, 2995.
13. Hildebrandt, P.; Stockburger, M. *Biochemistry* **1984**, 23, 5539.
14. Marcus, M.A.; Lewis, A. *Science* **1977**, 195, 1328.
15. Braiman, M.; Mathies, R. *Biochemistry* **1980**, 19, 5421.
16. El-Sayed, M.A.; Hsieh, C.-L. *Time-Resolved Vibrational Spectroscopy* **1983**.
17. Campion, A.; Turner, J.; El-Sayed, M.A. *Nature* **1977**, 265, 659.
18. Hsieh, C. L.; Nagumo, M.; Nicol, M.; El-Sayed, M. A. *Journal of Physical Chemistry* **1981**, 85, 2714.
19. Hsieh, C.L.; El-Sayed, M.A.; Nicol, M.; Nagumo, M.; Lee, J.H.
20. *Photochemistry and Photobiology* **1983**, 38, 83.
21. Vandenberg, R.; Jang, D.J.; Bitting, H. C.; El-Sayed, M.A. *Biophysical Journal* **1990**, 58, 135.
22. Griffiths, J. A.; Masciangioli, T. M.; Roselli, C.; El-Sayed, M. A. *Journal of Physical Chemistry* **1996**, 100, 6863.

